

Antimicrobial Resistance and Toxin Gene Profiles of Clostridioides (Clostridium) difficile Isolates from Diverse Fecal Contaminated Environmental Sources

Antimicrobial Resistance and Toxin Gene Profiles of Clostridioides (Clostridium) difficile Isolates from Diverse Fecal Contaminated Environmental Sources
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Introduction
Clostridioides (ex former Clostridium) difficile is a spore forming Gram positive anaerobe which is considered as an important pathogen causing antibiotic-related associated intestinal disease in humans and some animal species, but can be present also in various environments outside the hospital. Little is known about the environmental isolates and few studies have been conducted on the prevalence and molecular epidemiology of C. difficile in local contaminated environmental samples.

Objectives of the study
The current study was done to investigate the prevalence and the molecular characteristics of toxin-producing genes and antimicrobial susceptibility profiles of such "non-toxin and C. difficile isolates in local contaminated environmental sources.

Materials and Methods
Sample collection: A total of 22 environmental samples (2 raw sewage, 2 treated sewage, 2 raw sludge, 2 digested sludge, 2 village (water or grass), fresh leaves, 12 solid waste, 2 others less than six months in age and 3 others more than six months in age) and 2 storage room samples were collected from wastewater treatment plant (WWT) and other sites. All samples were handled in the laboratory within 24 hours collection.
Fig. 1: Flow diagram of the experimental steps for isolation, identification, detection of enterotoxigenic genes and antimicrobial susceptibility of C. difficile from various environmental samples.

Results
Overall, C. difficile was isolated from 40.91% (9/22) of the samples (Table 1). C. difficile was isolated directly or after selective enrichment in raw sewage, 50.0% treated sewage effluents, 50% raw sludge, 50% digested sludge, 50% village water/grass, 50% fresh leaves (others less than six months in age), 55.56% fresh leaves (others more than six months in age), 6.67% and sludge, 0.0%. A total of 31 C. difficile isolates were isolated, and among them, 58.77% (18/31) of the isolates were non-toxinogenic, 32.26% (10/31) were A, B, CDT and 9.87% (3/31) were binary toxin-producers (A-B-CDT+).

Table 1: Prevalence, toxin-producing genes and antimicrobial resistance profiles of C. difficile in local contaminated environmental samples.

Sample	Isolation	Toxin genes	Antimicrobial resistance
Raw sewage	2	1	1
Treated sewage	2	2	2
Raw sludge	2	2	2
Digested sludge	2	2	2
Village (water or grass)	2	2	2
Fresh leaves	12	6	6
Others less than six months in age	2	1	1
Others more than six months in age	3	1	1
Storage room	2	0	0
Total	31	18	18

Discussion and Conclusions
Overall, the most of C. difficile isolates were non-toxicogenic, except one isolate was toxicogenic.
Enrichment culture was significantly more successful at detecting C. difficile than direct plating.
Since all these isolates were isolated directly or after selective enrichment on C. difficile selective agar (CDTA) supplemented only with vancomycin, Clostridium spp. other than C. difficile grown on CDTA. For this reason, confirmation should be conducted with

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INTRODUCTION

Clostridioides (or former *Clostridium*) *difficile*, is a spore forming Gram positive anaerobe which considered as an important pathogen causing antimicrobial-associated intestinal disease in humans and some animal species, but can be present also in various environments outside the hospital. Little is known about the environmental strains and few studies have been conducted on the prevalence and molecular epidemiology of *C. difficile* in fecal contaminated environmental sources. Therefore, optimization methods for isolation and molecular epidemiology of *C. difficile* are required to elucidate the role of the environmental sources as transmission routes for human infection.

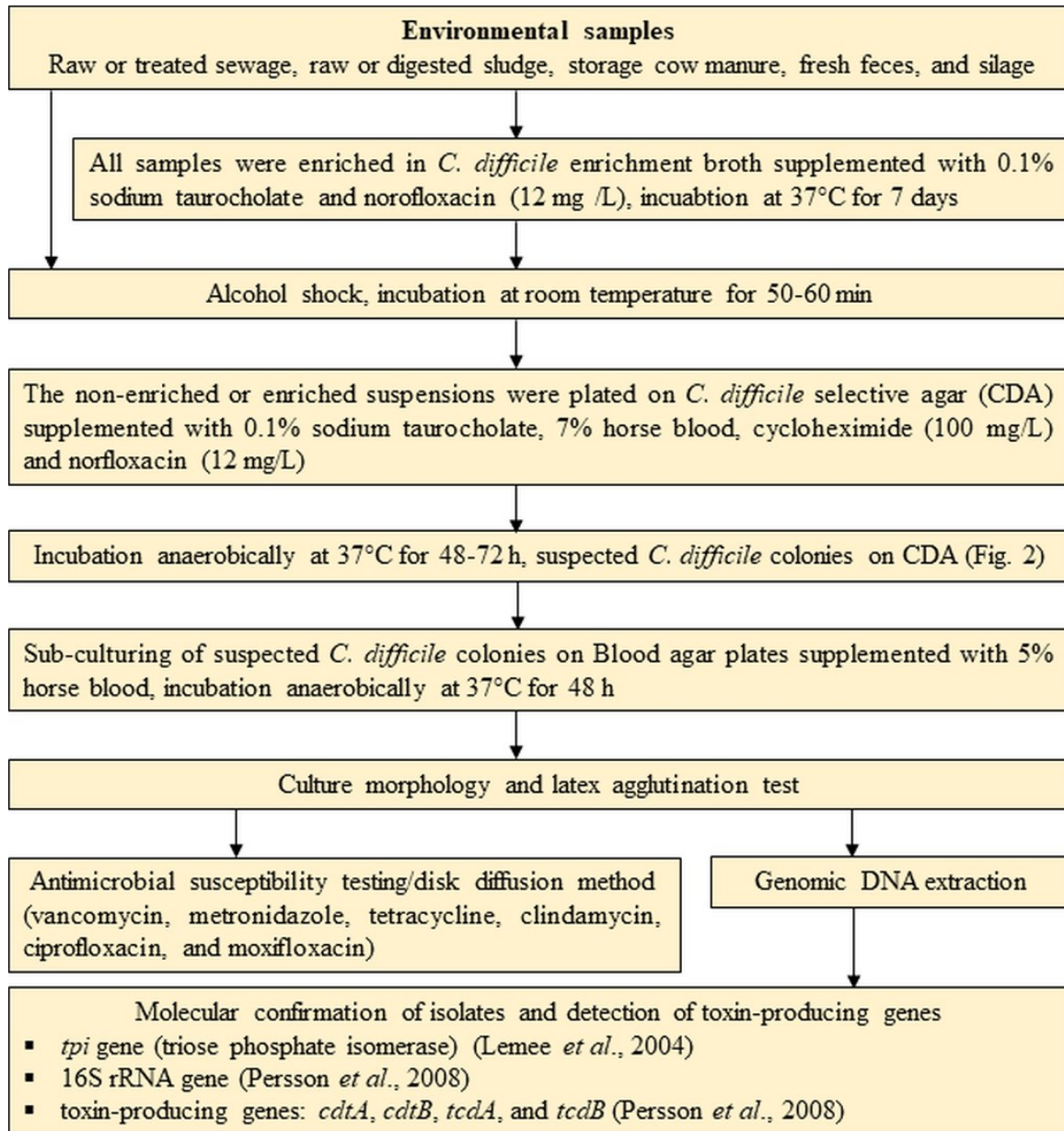
OBJECTIVES OF THE STUDY

The current study was done to investigate the presence and the molecular characteristics of toxin-producing genes and antimicrobial susceptibility profiles of such “environmental” *C. difficile* isolates in fecal contaminated environmental sources.

MATERIALS AND METHODS

Sample collection: A total of 22 environmental samples (2 raw sewage, 2 treated sewage, 2 raw sludge, 2 digested sludge, 2 silage (maize or grass), fresh feces (4 adult cows, 3 calves less than six months in age and 3 calves more than six months of age), and 2 storage cow manure) were collected from wastewater treatment plant (WWTP) and cattle farm. All samples were handled in the laboratory within 24 h after collection.

Fig 1: Flow diagram of the experimental setup for isolation, identification, detection of toxin-producing genes and antimicrobial susceptibility of *C. difficile* from various environmental samples.



RESULTS

- Overall, *C. difficile* was isolated from 40.91% (9/22) of the samples (Table 1).
- *C. difficile* was isolated directly or after selective enrichment in raw sewage, 100%; treated sewage effluents, 50%; raw sludge, 100%; digested sludge, 100%; storage cow manure, 50%; fresh feces (calves less than six months in age), 33.33%; fresh feces (adult cows), 0.0%; fresh feces (calves more than six months of age), 0.0% and silage, 0.0%.
- A total of 31 *C. difficile* isolates were isolated, and among them, 96.77% (30/31) of the isolates were non-toxigenic, 3.23% (1/31) was A⁻B⁺CDT⁻ and 6.45% (2/31) were binary toxin-positive (A⁻B⁻CDT⁺).

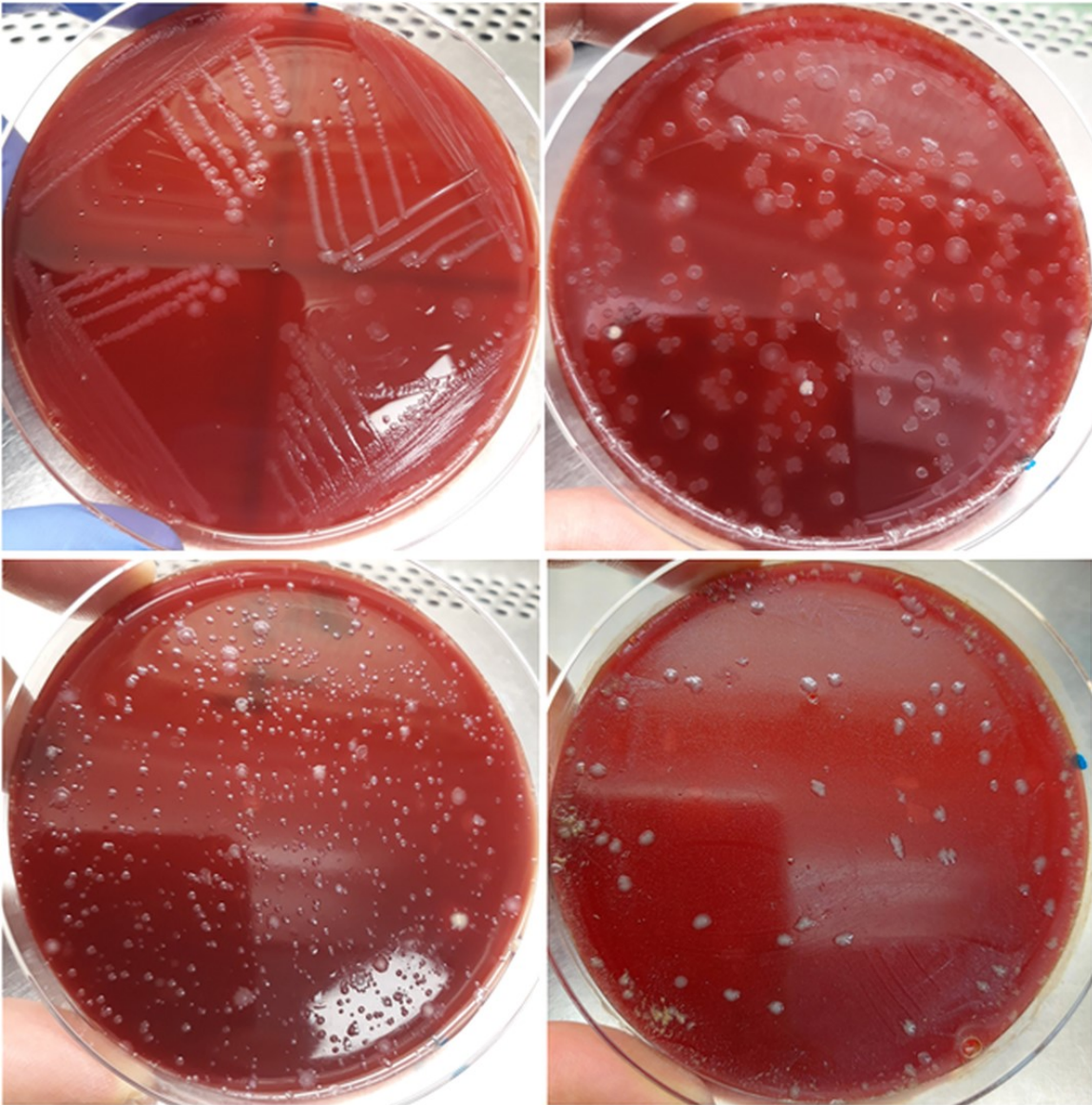
Table 1: Prevalence, toxin-producing genes and antimicrobial resistance profiles of *C. difficile* in fecal contaminated environmental samples.

Isolate No.	Sample type	Prevalence (%)	Isolation method	<i>tpi</i> gene	Toxin genes		Antimicrobial resistance profiles
					<i>tcdA</i> , <i>tcdB</i> , <i>cdtA</i> , <i>cdtB</i>		
RSD-D4	Raw sludge	2/2 (100%)	Direct	+	<i>tcdB</i>	-	
RSD-D7			Direct	+	<i>cdtB</i>	CIP	
RSD-D8			Direct	+	<i>cdtB</i>	CIP	
RSD-D9			Direct	+		CIP	
RSD-E1			Enrichment	+		CIP, VAN, CLI, MXF	
RSD-E2			Enrichment	+		VAN, CIP	
RSD-E6			Enrichment	+		VAN, CIP, MXF, TE	
RSD-E8			Enrichment	+		CIP, VAN	
RSD-E9			Enrichment	+		VAN, CIP	
RSD-E15			Enrichment	+		MTZ, CLI, CIP	
DSD-D5	Digested sludge	2/2 (100%)	Direct	+		VAN	
DSD-D22			Direct	+		VAN, CIP, MTZ	
DSD-E10			Enrichment	+		VAN, CIP, CLI	
RS-E1	Raw sewage	2/2 (100%)	Enrichment	+		VAN, CIP	
RS-E2			Enrichment	+		VAN, CIP	
RS-E4			Enrichment	+		VAN, CIP	
RS-E11			Enrichment	+		VAN, CIP, CLI, TE	
RS-E15			Enrichment	+		VAN, CIP, CLI, TE	
TS-E1			Treated sewage	1/2 (50%)	Enrichment	+	
TS-E2	Enrichment	+				VAN, CIP, TE	
TS-E3	Enrichment	+				VAN, CIP, TE	
TS-E4	Enrichment	+				VAN, CIP, TE	
TS-E5	Enrichment	+				VAN, CIP, TE	
TS-E6	Enrichment	+				VAN, CIP, TE	
TS-E7	Enrichment	+				VAN, CIP, TE	
TS-E8	Enrichment	+				VAN, CIP, TE	
TS-E10	Enrichment	+				VAN, CIP, TE	
CD3.1.1D	Fresh feces (calves less than six months age)	1/3 (33.3%)			Direct	+	
CD3.1.2D			Direct	+		MTZ, CIP, VAN	
CD3.1.3D			Direct	+		MTZ, CIP, VAN	
SM1-D	Storage cow manure	1/2 (50%)	Direct	+		MTZ, CIP, VAN	
nd.	Silage (grass or maize)	0/2 (0.0%)					
nd.	Fresh feces (calves more than six months age)	0/3 (0.0%)					
nd.	Fresh feces (adult cows)	0/3 (0.0%)					

VAN: vancomycin, CIP: ciprofloxacin, TE: tetracycline, CLI: clindamycin, MXF: moxifloxacin, MTZ: metronidazole. nd: not detected. Interpretation of inhibition zone: breakpoints of VAN, MXF and MTZ (Erikstrup *et al.*, 2012); Breakpoints of TE and CIP (Kouassi *et al.*, 2014); CLI breakpoint as recommended by members of the SFM Antibiogram Committee (2020).

- One toxigenic isolate (A⁻B⁺CDT⁻) was susceptible to all tested antimicrobials; while most non-toxigenic isolates were resistant to more than two antimicrobials of different classes. The resistance against fluoroquinolones (ciprofloxacin) is very common (93.55% of the isolates tested), glycopeptide antimicrobials (vancomycin, 83.87%), and tetracyclines (tetracycline, 38.71%), followed by lincosamide (clindamycin) and metronidazole (nitroimidazoles).

Fig. 2: Suspected *C. difficile* colonies (grey-white and irregular) on CDA plates.



DISCUSSION AND CONCLUSIONS

- Overall, the most of *C. difficile* isolates were non-toxigenic, except one isolate was toxigenic.
- Enrichment culture was significantly more successful at detecting *C. difficile* than direct plating.
- Since all these isolates were isolated directly or after selective enrichment on *C. difficile* selective agar (CDA) supplemented only with norfloxacin, *Clostridium* spp. other than *C. difficile* grew on CDA. For this reason, moxalactam should be combined with norfloxacin at concentration 32 mg/L as this inhibited the growth of *Clostridium* spp. (other than *C. difficile*).
- The *C. difficile* was found in fresh feces of calves less than six months in age, but not in other fecal samples (adult cows and calves more than six months of age), which considered to be a major reservoir of *C. difficile*.
- The majority of non-toxigenic isolates were found to be resistant to ciprofloxacin, vancomycin and tetracycline. These isolates could be carried tetracycline resistance genes [such as *tet*(M) and *tet*(W)] and macrolide-lincosamide-streptogramin B (MLS_B) resistance genes (*ermB*) often located on the mobile genetic elements (MGEs) such as conjugative transposons (Peng *et al.*, 2017).
- *C. difficile* strains might be acquired resistance to tetracycline and clindamycin via the transfer of MGEs among *C. difficile* strains and/or between *C. difficile* and the other bacterial species, especially conjugative transposons (Tn5397 and Tn916 which associated with tetracycline resistance, *tet*(M) and Tn5398, Tn6194 and Tn6215 associated with MLS_B family, *ermB* (Peng *et al.*, 2017).
- *C. difficile* strains are commonly present in various fecal contaminated environmental sources, which could be serve as a potential source of community-associated *C. difficile* infection.
- Molecular epidemiology is needed to determine the clinical relevance of environmental *C. difficile* strains isolated from environmental sources.

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